Effect of Peripheral 5-HT on Glucose and Lipid Metabolism in Wether Sheep

by Dian Wahyu Harjanti

Submission date: 07-Jan-2020 04:06PM (UTC+0700)

Submission ID: 1239735832

File name: se and Lipid Metabolism in Wether Sheep Dian W Harjanti 2014.pdf (1.35M)

Word count: 7606

Character count: 41092



Effect of Peripheral 5-HT on Glucose and Lipid Metabolism in Wether Sheep

Hitoshi Watanabe¹, Ryo Saito¹, Tatsuya Nakano¹, Hideyuki Takahashi¹, Yu Takahashi¹, Keisuke Sumiyoshi¹, Katsuyoshi Sato¹, Xiangning Chen¹, Natsumi Okada¹, Shunsuke Iwasaki¹, Dian W. Harjanti², Natsumi Sekiguchi², Hiroaki Sano², Haruki Kitazawa³, Michael T. Rose⁴, Shyuichi Ohwada¹, Kouichi Watanabe¹, Hisashi Aso¹*

1 Cellar Biology Laboratory, Graduate School of Agricultural Science, Tohoku 27 ersity, Sendai, Japan, 2 Department of Animal Sciences, Faculty of Agriculture, Iwate University, Morioka, Japan, 3 Laboratory of Food and Biomolecular Science, Graduate School of Agricultural Science, Tohoku University, Sendai, Japan, 4 Institute of Biological, Environmental and Rural Sciences, Aberystwyth University, Cardiganshire, United Kingdom

Abstract

In mice, peripheral 5-HT induces an increase in the plasma concentrations of glucose, insulin and bile acids, and a decrease in plasma triglyceride, NEFA and cholesterol concentrations. However, given the unique characteristics of the metabolism of rumina 10 relative to monogastric animals, the physiological role of peripheral 5-HT on glucose and lipid metabolism in sheep remains to be established. Therefore, in this study, we investigated the effect of 5-HT on the circulating concentrations of metabolites and insulin using five 5-HT receptor (5HTR) antagonists in sheep. After fasting for 24 h, sheep were intravenously injected with 5-HT, following which-, plasma glucose, insulin, triglyceride and NEFA concentrations were significantly elevated. In contrast, 5-HT did not affect the plasma cholesterol concentration, an 78 induced a decrease in bile acid concentrations. Increases in plasma glucose and insulin concentrations induced by 5-HT were attenuated by pretreatment with 9 lethysergide, a 5HTR 1, 2 and 7 antagonist. Additionally, decreased plasma bile acid concentrations induced by 5-HT were blocked by pre-treatment with Ketanserin, a 5HTR 2A antagonist. However, none of the 5HTR antagonists inhibited the increase in plasma triglyceride and NEFA levels induced by 5-HT. On the other hand, mRNA expressions of 5HTR1D and 1E were observed in the liver, pancreas and skeletal muscle. These results suggest that there are a number of differences in the physiological functions of peripheral 5-HT with respect to lipid metabolism between mice and sheep, though its effect on glucose metabolism appears to be similar between these species.

Citation: Watanal 23 Saito R, Nakano T, Takahashi H, Takahashi Y, et al. (2014) Effect of Peripheral 5-HT on Glucose and Lipid Metabolism in Wether Sheep. PLoS ONE 9(2): e88058. doi:10.1371/journal.pone.0088058

Editor: Hubert Vau University of Rouen, France, France

Received April 30, 2013; Accepted January 6, 2014; Published February 4, 2014

Copyright: © 2014 Watanabe et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any merita m, provided the original author and source are credited.

Funding: This research was supported by grants from the Ministry of Agricul 19; Forestry and Fisheries (2004), the Research Project on Development of Agricultural Products and Foods with Health-promoting benefits (NARO) and the Ministry of Edu 3 ion, Culture, Sports, Science and Technology, Japan Society for the Promotion of Science (JSPS), Research Fellowship for Young Scientists Program, Japan. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: asosan@bios.tohoku.ac.jp

Introduction

Serotonin (5-HT) is a monoaminers 10 heurotransmitter in the nervous system. In the periphery, 5 26 is produced from intestinal enterochromaffin cells by tryptophan hydroxylase (TPH) I, which is the rate-limiting enzyme for 5-HT biosynthesis in this tissue. Another isoform of this enzyme is TPH2, which is found in the central n 31 us system [1-2]. Peripheral 5-HT acts as a hormone, affecting vasoconstriction, intestinal motility, primary haemostasis, liver repair and the control of T cell-mediated immunity [3-8]. The periphery contains approximately 98% of the body's 5-HT. Additionally, 5-HT is thought not to be able to pass the blood-brain barrier [9-10]. Thus, there are two independ 43 5-HT synthesis systems: one in the periphery and the other in the central nervous system.

It has been reported that increasing c a plating concentrations of 5-HT induces hyperglycemia through the release of adrenaline from the adrenal gland in rats via action through the 5HTR1A, 2A or 7 [11–13]. Additionally, 5-HT stimulates glycogen synthesis

at nanomolar concentrations but inhibits it at micromolar concentrations in hepatocytes by serotonergic mechanisms [14], and 5-HT enhances net hepatic glucose upt 73 under hyperglycemic and hyperinsulinemic conditions [15]. On the other hand, we have previously reported that peripheral 5-HT decreases 6 sma lipid levels via action through several 5HTRs in mice [16]. These results suggest that peripheral serotonin plays an important role in glucose and lipid metabolism.

It is well established that glucose and lipid metabolism in ruminants are substantially different to those of humans and rodents. For example, ruminants again rb little glucose and have no glucokinase activity in the liver, and nearly all of their glucose to their glucose compensated for by gluconeogenesis [17–19]. Propionate is the principal source of carbon for glucose synthesis in liver, which meets 85–90% of the body glucose requirements in sheep [17,20]. Sheep are also more resistant to insulin action compared with non-ruminants [18,21]. Additionally, it has been shown that glucose, acetate, lactate and pyruvate are kage away materials in fatty acid biosynthesis in ruminants, and the rate of

fatty acid synthesis from glucose in ruminant adipose tissue is very low [22–25]. Furthermore, some reports have suggested that there are differences in secretion and function of gastric hormones between monogastric species and ruminants. For instance, glucose is able to inhibit the secretion of ghrelin in non-ruminants, though short chain fatty acids decrease plasma ghrelin concentrations in wethers [26–28]. Ghrelin injection also increases plasma glucose concentrations in adult cows, while no hyperglycemic response to ghrelin is o accepted an acid suckling or pre-ruminant calves [29]. In non-ruminant and se salts, amines, tastants and olfactants regulate 5-HT secretion from the enterochromaffin cells of the gastrointestinal tract [30–34]. Hence, it is likely that not only the secretion of 5-HT but also the response to 5-HT in ruminants may differ from that of monogastric animals, following the absorption of the products of rumen fermentation.

There are few re 42 s of the effects of peripheral 5-HT in ruminant animals on glucose and lipid metabolism, even though it has been show 1 to be involved in those of mice. In the present study, in order to clarify the distinctive features of peri 72 al 5-HT in glucose and lipid metabolism of sheep, we explored the effect of 5-HT on the concentrations of plasma metabolites and insulin.

Materials and Methods

Ethics statement

The experiments were permitted by the Tohoku University and to University Environmental & Safety Committee and conducted in accordance with the Guidelines for Animals Experimentation of Tohoku University and Iwate University, which have been sanctioned by the relevant committee of the Government of Japan based on the Declaration of Helsinki.

Experimental animals

Six, crossbred (Corriedale × Suffolk) wethers, 1–3 years of age, weighing 47.2±7.8 kg were used for this study. Three animals were chosen at random treatment experiments. Animals were fed a maintenance diet of alfalfa hay cubes, and mixed orchard grass and reed canary grass hay at 1300 h each day. Water was available ad libitum throughout the experiment. In order to stably maintain the concentrations of plasma metabolites during the injection experiments, the animals were used for the experiments after fasting for 24 h. The weight of each wether did not change during the study. Experiments were then conducted at weekly intervals. A catheter was inserted into the jugular vein of the animals and was filled with sodium citrate (0.13 mol/l) at 0900 h of the study day, and the injection study was performed at 1300 h.

5-HT and 5-HT receptor antagonist treatment experimen 34

5-HT (40 µg/kg body weight) (Sigma Chemical Co., St. Louis, MO) was dissolved in 5 ml phosphate buffered saline (PBS). Six animals were injected with a single dose of 5-HT into the jugular catheter. The dose used was determined according to previously published reports [35–37]. To eliminate the possibility of contamination by 5-HT in the catheter line, 10 ml PBS was flushed into the catheter 69 r 5-HT injection and 5 ml of blood was disposed of prior to sampling. Blood samples were collected from the jugular catheter every 15 min from –30 to –15 min, every 3 min from 0 to 15 min, every 15 min from 30 to 60 min, every 30 min from 90 to 120 min, and every 60 min from 180 to 360 min, relative to the injection of 5-HT. Two ml of blood was obtained at each sampling time point. After a week, three of the experimental animals were used for the 5HTR antagonist pre-

treatment experiments. Methysergide (antagonist for 5HTR1, 2 and 7; 40 µg/kg body weight; Sigma), SB-269970 (antagonist for 5HTR7; 70 µg/kg body weight; Tocris Bioscience, Ellisville, MO), SB-204070 (antagonist for 5HTR4; 40 µg/54 body weight; Sigma) and Ro 04-6790 (antagonist for 5HTR6; 30 µg/kg body weight; Sigma) were dissol 53 in PBS, respectively. Ketanserin (antagonist for 5HTR2A; 10 µg/kg body weight; Sigma) was dissolved in 0.1 M HCl, diluted with PBS. The final injection volume of all antagonists administrated was 5 ml. The doses of 5HTR antagonists were determined according to several published reports, bearing in mind the do 16 f 5-HT used [13,16,38-39]. These were administered 15 min prior to the injection of 5-HT. In addition the blood sampling times for the 5-HT injection noted above, blood samples were also obtained from the jugular vein at 10 min and 5 min before the 5-HT injection for the antagonist experiments. The animals were also treated with PBS as control, and blood samples were taken between -30 52 d 120 minutes. Blood samples for the 5-HT assay were placed into a polyethylene tube containing EDTA (7 ng/ml) and centrifuged at 4500×g for 10 min at 4°C. Samples for the other analyses were placed into a polyethylene tube containing h 7 arin (10 U/ml) and centrifuged at 9000×g for 10 min at 4°C. All samples 35 re kept on ice until centrifugation. Following centrifugation, plasma was harvested and stored at -20°C until analysis.

Biochemical analysis

The plasma 5-HT and insulin concentrations were determined by commercial ELISA kit, which were purchased from Immunotech (Marseille, 51 nce) and Mercodia (Uppsala, Sweden), respectively. The plasma concentrations of glucose, TG, NEFA, cholesterol and bile acids were measured using a commercial colormetri 6 kits (Wako, Osaka, Japan). All procedures were performed according to the respective manufacturer's instructions.

Total RNA preparation and quantitative real-time PCR analysis

Total RNA were extracted from liver, pancreas, semiter 33 osus muscle and vastus intermedius muscle using Isogen II reagent according to the manufacturer's instructions (NIPPON GENE Inc., Tokyo, Japan). cDNA templates were synthesized with the Superscript III RT kit (Invitrogen, Co., Carlsabad, 32) using random primers. The quantitative real-time PCR was performed in duplicate using the Thermal Cycler Dice Real Time System Single (Takara Bio Inc., Siga, Japan) to assay specific 5HTR1A, 5HTR1B, 5HTR1D, 5HTR1E, 5HTR1F, 5HTR2A, 5HTR2B, 5HTR2C, 5HTR3A, 5HTR3B, 5HTR4, 5HTR5A and 5HTR7 mRNA levels. The sequences of primers and sizes of each PCR product are listed in Table 1. The typi 13 reaction cycles consisted of an initial denaturation step 95°C for 30 sec followed by 40 cycles of denaturation at 95°C for 5 sec a 30 annealing at 60 to 64°C for 30 sec. The resolution curve was measured at 95°C for 15 sec, 60°C for 15 sec and 95°C for 15 sec. Quantification of the RT-PCR products was normalized to the endogenous housekeeping gene (18s) expression.

Statistical analysis

Values are reported as means \pm S.D. Average concentrations of metabolites and hormones from -30 to 0 min before the 5-HT injection were taken to be basal values. Statistical significances between basal and subsequent conce 41 ations of hormones and metabolites were determined using paired Student's t-test. To assess the effect of 5HTR antagonists, the incremental area was calculated by trapezoidal integration. Statistical significances

Table 1. Primers used in quantitative real-time PCR analysis.

Genes		Primer sequence (5'-3')	Product size (bp) Tm °		
5HTR 1A	Forward	AATGGCCGCGTTGTACCAG	145	64	
	Reverse	GGTGATGGCCCAGTATCTATCCAG			
5HTR 1B	Forward	TGATGCCCATCAGTACCATGT	71	60	
	Reverse	CCAGAAGTCGCAGACCACCT			
5HTR 1D	Forward	ACCGCGCATCTCATCACAG	142	64	
	Reverse	GTTCCAGGACACTATCGGCAAG			
5HTR 1E	Forward	AGTGTGGCTGTGAGACCCAAGA	105	64	
	Reverse	CACGATCACGGCGGAGTTTA			
5HTR 1F	Forward	CAAGCAAGTAGGATTGCCAAGGA	101	64	
	Reverse	TTTGCTAGCATGTACGGTGTGGA			
5HTR 2A	Forward	CCAGCCTTGGCCTACAAGTC	84	64	
	Reverse	GTCATTATCTGTCGTCTTGCCATC			
5HTR 2B	Forward	GCCTCACCTACAGACATGGACAGA	117	64	
	Reverse	AGGCTTTGTACCCATGCCAAAC			
5HTR 2C	Forward	GATTTGAACCCACGCCGAAG	134	64	
	Reverse	AATGGGCACCACATGATCAGAA			
5HTR 3A	Forward	CACCTGCTGGCCAACTACAAGAA	101	64	
	Reverse	ACGCTGAGGATGGCATAGACAA			
5HTR 3B	Forward	TGCAGAACAGCGCTGGAGA	83	64	
	Reverse	CAGGCTCACCACGTAGACCAGA			
5HTR 4	Forward	CCTTGAATCTGGCCTTGCTG	117	64	
	Reverse	CTTGAGCACTGCTTGGTCCTG			
5HTR 5A	Forward	GAAGATCTACAAGGCCGCCAAG	74	64	
	Reverse	CGGTTTCGGATATGGGTGAGAC			
5HTR 7	Forward	CACTGCGGTAAGCCTAGTGATGAA	106	64	
	Reverse	GCTTTGAACGGACACTGCTCTG			
18s	Forward	GCCCTATCAACTTTCGATGGTAGTC	113	64	
	Reverse	CCTTGGATGTGGTAGCCGTTTC			

doi:10.1371/journal.pone.0088058.t001

between the PBS treatment and the antagonist treatm 68 were determined using one-way ANOVA. P values less than 0.05 were considered to be statistically significant.

Results

Plasma 5-HT concentrations after injection of 5-HT in sheep

Relative to an intravenous injection of 5-HT, the plasma concentrations of 5-HT were measured between $-30~\rm min$ and 360 min (Figure 1). The average basal concentration $40~\rm lasma$ 5-HT was approximately 0.1 μM before the injection of 5-HT. After the 5-HT injection, the plasma concentration of 5-HT reached a peak of 0.6 μM at 3 min, and sharply decreased thereafter. Concentrations returned to basal levels by 60 min.

Effect of peripheral 5-HT on plasma glucose concentrations in sheep

Following 5-HT injection, the plasma glucose cond 67 ation was significantly elevated from 3 min to 45 min, reached a peak at 6 min, gradually returned to basal levels, and was slightly lower than basal levels between 240 min to 360 min (Figure 2a). No

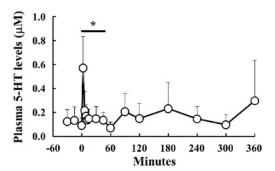


Figure 1. Plasma concentration of 5-HT following 5-HT injection. Blood samples were obtained from 6 sheep between -30 and 360 min region were obtained injection of 5-HT (40 μ g/kg body weight). The concentration of plasma 5-HT was measured. *: 9<0.05 relative to the basal average values from -30 min to 0 min. doi:10.1371/journal.pone.0088058.g001

significant variations in circulating glucose concentrations were observed before the 5-HT injection. In order to determine what kind of 5HTRs were related to the hyperglycemia induced by 5-HT, sheep were pre-treated with several 5HTR antagonists: Methysergide (5HTR1, 2 and 7), Ketanserin (5HTR2A), SB-269970 (5HTR7), SB-204070 (5HTR4) or Ro 04-6790 (5HTR6) at 15 min before 5-HT injection. Pre-treatments of all 5HTR antagonists did not influence the concentration of plasma metabolites. Pre-treatment with Methysergide completely inhibited the hyperglycemia induced by 5-HT. However, pretreatment with the other antagonists did not significantly affect hyperglycemia. Additionally, plasma glucose concentrations were not affected between -30 and 120 min by the injection of PBS (Figure 2b). The incremental areas of glucose concentrations following 5-HT injection alone and following pretreatment with Ketanserin, SB-269970, SB-204070 and Ro 04-6790 were significantly greater than that of PBS injection (Figure 2c). In contrast, the incremental area following pre-treatment with Methysergide was not significantly increased compared with that of PBS injection. These data suggest that peripheral 5-HT increases plasma glucose concentrations through 5HTR1.

Effect of peripheral 5-HT on plasma insulin concentrations in sheep

After the 5-HT injection alone, plasma insulin concentrations were significantly increased between 6 and 15 min post injection. Levels were maximal at 9 min and were not significantly different from basal values after 30 min (Figure 3a). The concentration of plasma insulin before the injection of 5-HT 50's constant. Pretreatment with Methysergide inhibited the increase in plasma insulin levels caused by administration of 5-HT in sheep, and levels were significantly lower than basal values between 120 and 180 min after the 5-HT injection. Pre-injection with Ketanserin, SB-204070 and Ro 04-6790 did not block the elevation in plasma insulin concentrations induced by 5-HT. However, in sheep pretreated with 66 269970, hypoinsulinemia was observed between 60 and 180 min after the 5-HT administration. On the other hand, the administration of PBS did not affect plasma insulin concentrations (Figure 3b). The incremental area for Methysergide was not significantly different to that for the PBS injection (Figure 3c). In contrast, insulin levels for the other antagonist pretreatments were significantly increased compared with that for PBS injection. These results suggest that peripheral 5-HT induces hyperinsulinemia through the 5HTR1.

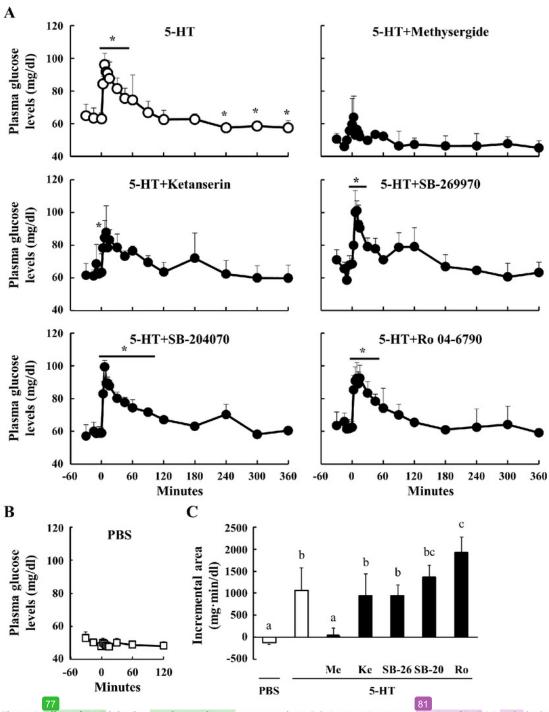


Figure 2. Effect of 5-HT injection on plasma glucose concentrations. Relative to an intravenous injection of 5-HT (40 μ g/kg body weight) at 0 min, plasma samples were obtained from 6 sheep between -30 and 360 min (n = 6). Three sheep were injected through the jugular catheter with several 5HTR antagonists (n = 3): Methysergide (antagonist for 5HTR1, 2 and 7, 40 μ g/kg body weight), Ketanserin (5HTR2A, 10 μ g/kg body weight), SB-269970 (5HTR7, 70 μ g/kg body weight), SB-204070 (5HTR4, 40 μ g/kg body weight), and Ro 04-6790 (5HTR6, 30 μ g/kg body weight), at 15 min before the injection of 5-HT. Plasma glucose concentrations were determined between -30 and 360 min (A). Plasma glucose levels after the injection of PBS were measured between -30 and 120 min $\frac{1}{5} = 3$) (B). The incremental area between 0 and 60 min was calculated (C). *: P<0.05 relative to the basal average values from -30 to 0 min. Columns with a different letter are significantly different (P<0.05). Me: Methysergide; Ke: Ketanserin; SB-26: 39 69970; SB-20: SB-204070; Ro: Ro 04-6790. doi:10.1371/journal.pone.0088058.g002

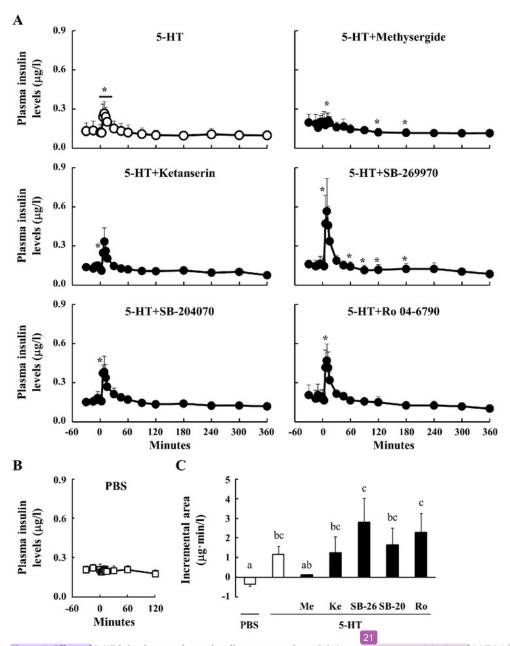


Figure 3. Effect of 5-HT injection on plasma insulin concentrations. Relative to an intravenous injection of 5-HT (40 μg/kg body weight) at 0 min, plasma samples were obtained from 6 sheep between -30 and 360 min (n=6). Three sheep were injected through the jugular vein with several 5HTR antagonists (n=3): Methysergide (antagonist for 5HTR1, 2 and 7, 40 μg/kg body weight), Ketanserin (5HTR2A, 10 μg/kg body weight), SB-269970 (5HTR7, 70 μg/kg body weight), SB-204070 (5HTR4, 40 μg/kg body weight), and Ro 04-6790 (5HTR6, 30 μg/kg body weight), at 15 min before the injection of 5-HT. Plasma insulin concentrations were determined between -30 and 360 min (A). Plasma insulin levels after the injection of PBS were measured between -30 and 120 min (n 5) (B). The incremental area between 3 and 15 min was calculated (C). *: P<0.05 relative to the basal average values from -30 to 0 min. Columns with a different letter are significantly different (P<0.05). Me: Methysergide; Ke: Ketanserin; SB-26: SB-204070; Ro: Ro 04-6790. doi:10.1371/journal.pone.0088058.g003

Effect of peripheral 5-HT on the plasma triglyceride concentrations in sheep

5-HT significantly induced an elevation in plasma triglyceride levels between 3 and $6\,\mathrm{min}$ after the injection. However,

concentrations were significantly decreased relative to the administration of 5-HT, plasma triglyceride levels were stable. Pre-injection with Methysergide, Ketanserin and SB-269970 did

not affect the hypertriglyceridemia induced by 5-HT, but they did appear to prevent the significant reduction in plasma triglyceride concentrations observed at 45 min after the 5-HT injection. Pretreatment with SB-204070 and Ro 04-6790 did not affect the profile of plasma triglycerides relative to 5-HT alone. Additionally, no significant effects of the PBS injection on plasma triglyceride concentrations were observed (Figure 4b). The incremental area of the PBS treatment (Figure 4c). These results indicate that the hypertriglyceridemia induced by 5-HT does not depend on 5HTR1, 2, 4, 6 or 7, and that 5HTR1, 2 and 7 may be related to the hypotriglyceridemia induced by peripheral 5-HT.

Effect of peripheral 5-HT on plasma NEFA and cholesterol concentrations in sheep

Plasma NEFA concentrations were significantly increased between 6 and 15 min after the 5-HT injection, but were decreased by 45 min, relative to the baseline levels (Figure 5a). The concentration of plasma NEFA was constant before the injection of 5-HT. Pre-administration with the antagonists did not attenuate the elevation of plasma NEFA concentrations in the minutes immediately following the 5-HT injection. However, pre-treatment with Methysergide and Ketanserin prevented the reduction in plasma NEFA concentrations at 45 min after 5-HT treatment. Additionally, the injection of PBS did not affect plasma NEFA concentrations (Figure 5b). Moreover, none of the 5HTR antagonists affected the incremental area under the NEFA concentrations induced by 5-HT (Figure 5c).

We have reported previously that peripheral 5-HT substantially decreases plasma cholesterol concentrations in mice [16]. However, 5-HT and PBS injections did not affect the plasma cholesterol concentration in sheep in the present experiment (Figure 6a, 6b). Additionally, none of the 5-HT antagonists affected the level of plasma cholesterol.

Effect of peripheral 5-HT on the concentration of plasma bile acids in sheep

The concentration of plasma bile acids was significantly lower following the 5-HT injection relative to basal levels. Couldn't rations returned to the baseline after 300 min (Figure 7a). 47 or to the injection of 5-HT, no significant differences in plasma bile acid concentrations were observed. The concentration of bile acids was decreased following 5-HT and pre-treatment with Methysergide, SB-269970, SB-204070 and Ro 04-6790. In contrast, the Ketanserin attenuated the reduction in the concentration of plasma bile acids induced by 5-HT. Additionally, plasma bile acid concentrations were not decreased by the administration of PBS (Figure 7b). Although the incremental area of bile acid concentrations following pre-treatment with Ketanserin was similar to that of PBS, those following 5-HT and pretreatment with the other antagonists were significantly lower than t of PBS (Figure 7c). These results suggest that 5-HT induces a decrease in the plasma concentrations of plasma bile acids through the 5HTR2A.

mRNA Expressions of 5HTRs in the liver, pancreas and skeletal muscle

To identify primary forms of 5HTRs in metabolically active tissues, we investigated the mRNA expression of 5HTRs in the liver, pancreas and skeletal muscle. In these tissues, 5HTR1D, 1E, 2B and 5A were the main forms expressed in these tissues; this was particularly the case for 5HTR1D (Figure 8). 5HTR4 was expressed only in the liver.

Discussion

There are two independent 5-HT systems, one in the brain and the other in the periphery. In humans and rodents, 5-HT regulates glucose and lipid metabolism through several 5HTRs and the 5-HT transporter (SERT) [62 6,40-41]. However, we are not aware of any reports on the function of pheral 5-HT in ruminant animals. The main objective of this study was to clarify the role of peripheral 5-HT in sheep by investigating the effect of intravenous 5-HT injection on the concentrations of metabolites related to glucose and lipid metabolism in the circulation.

5-HT is known to be associated wit 80 lucose metabolism, mainly because of its regulation of the secretion of insulin in pancreatic \$\beta\$ cells. The insulin secretion induced by glucose is inhibited by 5-HT in rat islet of Largerhans incubations in vitro [41]. Another report demonstrates that 5-HT regulates insulin secretion by serotonylation of GTPase within the pancreatic βcells [40]. Additionally, 5-HT directly controls the uptake of glucose into the 61 heral tissues, including the liver and skeletal muscles [42-43]. 5-HT enhances net hepatic glucose uptake under hyperglycemic and hyperinsulinemic conditions [15], and stimulates glycogen synthesis at nanomolar concentrations, but inhibits it at micromolar concentrations by serotonergic mechanisms in hepatocytes [14]. More 79 r, we previously reported that 5-HT induced the elevation of plasma glucose and insulin concentrations through different 5HTRs in mice, and that hyperglycemia after the injection of 5-HT was induced by repressing glucose uptake into the tissues [16].

Considering the data in this report, there appear to be a number of differences in 560 action between ruminants and nonruminants; the effect of 5-HT on plasma metabolite and insulin concentrations in sheep and mice is summarized in Table 2. In this study, plasma glucose and insulin concentrations were increased following the 5-HT injection, as seen in mice. Additionally, elevated plasma glucose and insulin levels in sheep were prevented following pre-treatment with Methysergide. In contrast, preinjection with Ketanserin and SB-269970 prevented the elevation of plasma glucose concentrations induced by 5-HT in mice, but not sheep. Moreover, high mRNA expressions of 5HTR1D and 1E were observed, but 5HTR2A and 7 mRNA expressions were lower in the liver and pancreas of sheep. These data indicate that intravenous 5-HT injection induces hyperglycemia and hyperinsulinemia only through the 5HTR1 in sheep. It is well established that the digestion, absorption, utilization and production of glucose in ruminants differ greatly from those of in monogastric animals [17-21]. However, the recovery to basal levels of plasma insulin after the 5-HT administration was 59 r than that of glucose in both sheep and mice, though an increase in plasma insulin concentrations was observed after the elevation of plasma glucose 58 following 5-HT injection [16]. In addition, 5-HT appears to d $_{57}$ ly regulate insulin secretion from pancreatic β cells [40-41]. Taken together, these data suggest that the elevation of plasma glucose and insulin concentrations induced by 5-HT may have independent pathways in sheep and mice.

In mice, plasma triglyceride, NEFA and cholesterol concentrations are decreased by peripheral 5-HT through 5HTR1, 2A and 7 (Table 2) [16]. In sheep, 5-HT administration resulted in an increase in plasma triglyceride and NEFA levels, but it did not affect plasma cholesterol levels in this study. Sheep are characterized by low concentrations of plasma VLDL and a high ratio of plasma HDL, compared with the mouse [44–46]. In addition, the concentration of each plasma lipid is also controlled by regulating the homeostatic uptake and release of lipids in the various tissues by 5-HT through several 5HTRs [47]. Therefore, the difference in

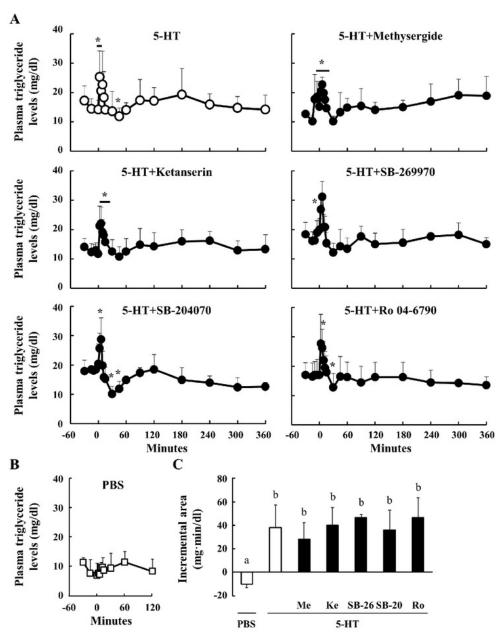


Figure 4. Effect of 5-HT injection on plasma triglyceride concentrations. Relative to an intravenous injection of 5-HT (40 μ g/kg body weight) at 0 min, plasma samples were obtained from 6 sheep between -30 and 360 min (n = 6). Three sheep were injected through the jugular vein with several 5HTR antagonists (n = 3): Methysergide (antagonist for 5HTR1, 2 and 7, 40 μ g/kg body weight), Ketanserin (5HTR2A, 10 μ g/kg body weight), SB-269970 (5HTR7, 70 μ g/kg body weight), SB-204070 (5HTR4, 40 μ g/kg body weight), and Ro 04-6790 (5HTR6, 30 μ g/kg body weight), at 15 min before the injection of 5-HT. Plasma triglyceride concentrations were determined between -30 and 360 min (A). Plasma triglyceride levels after the injection of PBS were measured between -30 and 120 min (n = $\frac{1}{5}$ B). The incremental area between 0 and 6 min was calculated (C). *: P<0.05 relative to the basal average values from -30 to 0 min. Columns with a different letter are significantly different (P<0.05). Me: Methysergide; Ke: Ketanserin; SB-26: SB-269970; SB-20: SB-204070; Ro: Ro 04-6790. doi:10.1371/journal.pone.0088058.g004

plasma lipid concentrations induced by peripheral 5-HT may be attributable to differences in the concentrations and the ratio of each lipoprotein between sheep and mice. Differences in their responses to 5-HT in terms of plasma lipid levels may be caused by different expressions of 5HTRs in the various tissues, as the metabolism and functions of each tissue in ruminant animals differ from those of in monogastric animals.

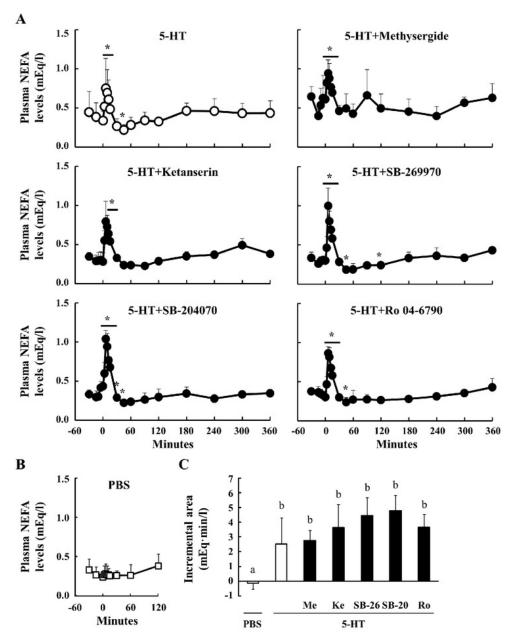


Figure 5. Effect of 5-HT injection on plasma NEFA concentrations. Relative to an intravenous injection of 5-HT (40 μ g/kg body weight) at 0 min, plasma samples were obtained from 6 sheep between -30 and 360 min (n = 6). Three sheep were injected through the jugular catheter with several 5HTR antagonists (n = 3): Methysergide (antagonist for 5HTR1, 2 and 7, 40 μ g/kg body weight), Ketanserin (5HTR2A, 10 μ g/kg body weight), SB-269970 (5HTR7, 70 μ g/kg body weight), SB-204070 (5HTR4, 40 μ g/kg body weight), and Ro 04-6790 (5HTR6, 30 μ g/kg body weight), at 15 min before the injection of 5-HT. Plasma NEFA concentrations were determined between -30 and 360 min (A). Plasma NEFA levels after the injection of PBS were measured between -30 and 120 min (n $\frac{53}{2}$) (B). The incremental area between 0 and 12 min was calculated (C). *: P<0.05 relative to the basal average values from -30 to 0 min. Columns with a different letter are significantly different (P<0.05). Me: Methysergide; Ke: Ketanserin; SB-26: SB-269970; SB-20: SB-204070; Ro: Ro 04-6790. doi:10.1371/journal.pone.0088058.g005

In mice, plasma triglyceride and NEFA concentrations were decreased by peripheral 5-HT injection; these effects v₁₈ blocked by SB-269970 and Ketanserin, respectively (Table 2). The present

study revealed that the effect of 5-HT on the plasma triglyceride and NEFA concentrations in sheep was opposite to that seen in mice. However, the decrease in plasma triglyceride and NEFA

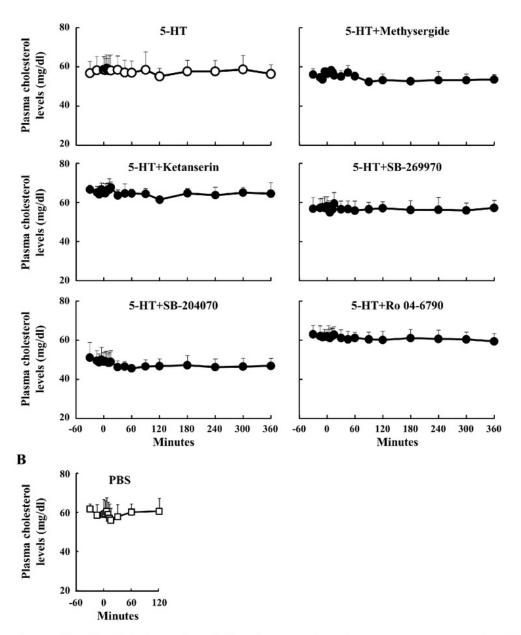


Figure 6. Effect of 5-HT injection on plasma cholesterol concentrations. Relative to an intravenous injection of 5-HT (40 μ g/kg body weight) at 0 min, plasma samples were obtained from 6 sheep between -30 and 360 min (n = 6). Three sheep were injected through the jugular catheter with several 5HTR antagonists (n = 3): Methysergide (antagonist for 5HTR1, 2 and 7, 40 μ g/kg body weight), Ketanserin (5HTR2A, 10 μ g/kg body weight), SB-269970 (5HTR7, 70 μ g/kg body weight), SB-204070 (5HTR4, 40 μ g/kg body weight), and Ro 04-6790 (5HTR6, 30 μ g/kg body weight), at 15 min before the injection of 5-HT. Plasma cholesterol concentrations were determined between -30 and 360 min (A). Plasma cholesterol levels after the injection of PBS were measured between -30 and 120 min (n = 3) (B). *: P<0.05 relative to basal average values from -30 min to 0 min. doi:10.1371/journal.pone.0088058.g006

concentrations relative to basal values was also observed in sheep, as for mice after each peak. The reduced plasma triglyceride level at 45 min after 5-HT injection in sheep was inhibited by three of the 5HTR antagonists: Methysergide, Ketanserin and SB-269970. In addition, decreased plasma NEFA concentrations at 45 min after injection of 5-HT was blocked by Methysergide and

Ketanserin. Thus, there is a similar inclination between sheep and mice in terms of the decrease of plasma triglyceride and NEFA concentrations induced by 5-HT, although not identical results of 5-HT treatment. On the other hand, 5-HT did not appear to affect the metabolism of cholesterol in sheep.

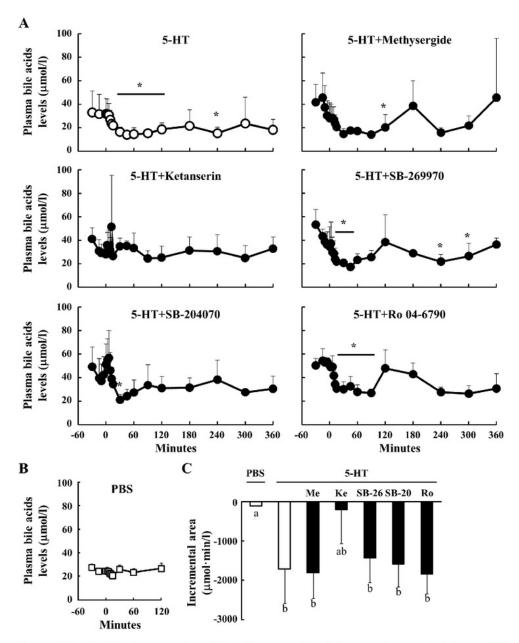


Figure 7. Effect of 5-HT injection on plasma bile acid concentrations. Relative to an intravenous injection of 5-HT (40 μ g/kg body weight) at 0 min, plasma samples were obtained from 6 sheep between -30 and 360 min (n=6). Three sheep were injected through the jugular vein with several 5HTR antagonists (n=3): Methysergide (antagonist for 5HTR1, 2 and 7, 40 μ g/kg body weight), Ketanserin (5HTR2A, 10 μ g/kg body weight), SB-269970 (5HTR7, 70 μ g/kg body weight), SB-204070 (5HTR4, 40 μ g/kg body weight), and Ro 04-6790 (5HTR6, 30 μ g/kg body weight), at 15 min before the injection of 5-HT. Plasma bile acid concentrations were determined between -30 and 360 min (A). Plasma bile acid levels after the injection of PBS were measured between -30 and 120 min (n=3) 15 The incremental area between 30 and 120 min was calculated (C). *: P<0.05 relative to the basal average values from -30 to 0 min. Columns with a different letter are significantly different (P<0.05). Me: Methysergide; Ke: Ketanserin; SB-26: SB-269970; SB-204070; Ro: Ro 04-6790. doi:10.1371/journal.pone.0088058.g007

After 5-HT injection, the concentration 12 plasma bile acids was decreased in sheep (Table 2). In contrast, the concentration of bile acids in plasma in mice is increased between 30 and 90 min after a

5-HT injection, due to an elevated re-absorption in the ileum [16] and an induced excretion from the gallbladder [48]. Additionally, after three days of ligation of the bile duct, mice lacking peripheral

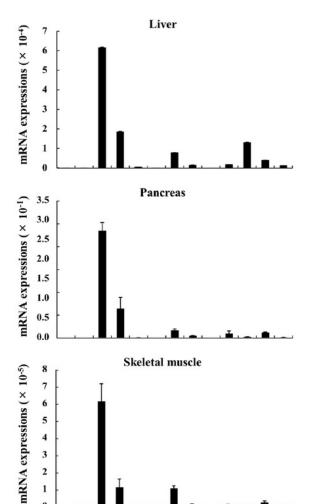


Figure 8. mRNA expressions of 5HTRs in the liver, pancreas and skeletal muscle. Liver, pancreas and skeletal muscle tissues were obtained from 3 sheep. After total RNA from e3 h tissue was extracted, 5HTRs mRNA expressions were measured by real-time PCR analysis. doi:10.1371/journal.pone.0088058.g008

1A 1B 1D 1E 1F 2A 2B 2C 3A 3B 4 5A

5-HT display higher levels of plasma bile salts than wild type mice, as Ost α and Ost β , the bile salt re-absorption transporters, are upregulated in the kidneys, along with a decrease in urinary bile salt excretion [49]. Besides this, in ruminants, the excretion of bile from the gallbladder occurs following the absorption of lipids [50]. Thus, rapid excretion of bile acids cannot occur following an increase in the plasma 5-HT concentration in sheep. Therefore, the decreased concentrations of plasma bile acids in sheep may be induced by 5-HT treatment without the excretion of bile from the gallbladder, in contrast to what is seen in rodents. In mice, Methysergide and Ketanserin antagonized the elevation in the concentration of plasma bile acids after 5-HT injection (Table 2). Additionally, Ketangain administration to sheep demonstrated that 5-HT induced a decrease in the concentration of plasma bile acids through 5H 72 A. These data indicate that 5-HT has the opposite effects on the concentration of plasma bile acids in sheep and mice, but through the same 5HTR2A. However, the expression of 5HTR2A was lower than other 5HTRs in the 17 ver of sheep. The non-hepatic tissues are probably concerned with a decrease in the concentration of bile acids in the circulation by 5-HT, though it is known that the liver is greatly concerned with bile acids met balism.

Fifteen 20 Γ receptors have been identified in the mouse. They have been divided into seven distinct classes, 5HTR1 to 7, largely on the 156 of their structure and operational characteristics. 5HTRs belong to the G-proteir 48 upled receptor (GPCR) subfamily, except for 5HTR3 28 ich is a ligand-gated ion channel, and all are found both i 381e central nervous system and in many peripheral tissues. 5-HT is thought to exert its effects through these membrane bound receptors [51]. However, there are 47 reports on the distribution of 5HTRs in sheep. Accordingly, in order to investigate the physiological role of 5-HT, the localization of 5HTRs in ruminants needs to be established.

Ruminants have a distinct metabolism with respect to their utilization and production of glucose and fatty acids [17-25]. Therefore, it is likely that 5-HT may also have different functions with regard to glucos and lipid metabolism in ruminants. Here, we have investigated the effects of 5-HT injection on glucose and lipid metabolisms in sheep. In conclusion, we note that peripheral 5-HT affects the metabolism of glucose in sheep through the same mechanism and 5HTRs as mice, but that it has different functions with respect to lipid metabolism between sheep and mice.

Table 2. Comparison of the effects of several 5-HT receptor antagonists on the plasma concentrations of metabolites and insulin, following 5-HT injection in sheep and mice.

	Sheep					Mouse ^a				
Plasma contents	5-HT ^b	Me	Ke	SB-26	SB-20	Ro	5-HT	Me	Ke	SB-26
Glucose	1	+	-	_	-	_	1	+	+	+
Insulin	1	+	-	-	-	-	1	+	-	-
Triglyceride	1	-	-	-	-	-	D	-	-	+
NEFA	1	-	-	-	-	-	D	-	+	-
Cholesterol	N	-	-	-	-	-	D	+	-	-
Bile acids	D	_	+	_	_	_	1	+	+	_

Murine data published previously [Ref. 16].

5-HT: serotonin; Me: methysergide; Ke: ketanserin; SB-26: SB-269970; SB-20: SB-204070; Ro: Ro 04-6790; I: increase; D: decrease; N: no change; +: effective in blocking ct of 5-HT; -: ineffective in blocking the effect of 5-HT. doi:10.1371/journal.pone.0088058.t002

1

Acknowledgments

We would like to express our thanks to the staff of the Department of Animal Sciences Laboratory, Iwate University, for maintaining the sheep and their excellent technical help.

References

- 1. Walther DJ, Peter JU, Bashamakh S, Hörtnagl H, Voits M, et al. (2003) Synthesis of serotonin by a second tryptophan hydroxylase isoform. Science 299:
- 2. Walther DJ, Bader M (2003) A unique central tryptophan hydroxylase isoform. Biochem Pharmacol 66: 1673-1680.
- 3. Chester AH, Martin GR, Bodelsson M, Arneklo-Nobin B, Tadjkarimi S, et al. (1990) 5-hydroxytryptamine receptor profile in healthy and diseased human epicardial coronary arteries. Cardiovasc Res 24: 932–937. 4. Geba GP, Ptak W, Anderson GM, Paliwal V, Ratzlaff RE, et al. (1996) Delayed-
- type hypersensitivity in mast cell-deficient mice: dependence on platelets for expression of contact sensitivity. J Immunol 157: 557–565.

 5. Hixson EJ, Lehrmann GV, Maickel RP (1977) Contractile responses to
- tryptamine analogues in isolated smooth muscle. Arch Int Pharmacodyn Ther
- 6. Holland JM (1976) Serotonin deficiency and prolonged bleeding in beige mice. Proc Soc Exp Biol Med 151: 32-39.
- 7. Lesurtel M, Graf R, Aleil B, Walther DJ, Tian Y, et al. (2006) Platelet-derived serotonin mediates liver regeneration. Science 312: 104-107.
- 8. Rapport MM, Green AA, Page IH (1948) Crystalline serotonin. Science 108:
- Merritt JH, Chamness AF, Allen SJ (1978) Studies on Blood-Brain Barrier Permeability After Microwave-Radiation. Radiat Environ Biophys 15: 367–377.
- 10. Woolley DW, Shaw E (1954) A biological and pharmacological suggestion about certain mental disorders. Proc Natl Acad Sci 40: 228–231.

 11. Chaouloff F, Laude D, Baudrie V (1990) Ganglionic transmission is a
- prerequisite for the adrenaline-releasing and hyperglycemic effects of 8-OH-DPAT. Eur J Pharmacol 185: 11–18.
- Sugimoto Y, Yamada J, Kimura I, Watanabe Y, Horisaka K (1992) The effects of the serotonin1A receptor agonist buspirone on the blood glucose and
- pancreatic hormones in rats. Jpn J Pharmacol 60: 145–148.

 13. Yamada J, Sugimoto Y, Yoshikawa T, Kimura I, Horisaka K (1995) The involvement of the peripheral 5-HT2A receptor in peripherally administered
- serotonin-induced hyperglycemia in rats. Life Sci 57: 819–825.

 14. Hampson IJ, Mackin P, Agius L (2007) Stimulation of glycogen synthesis and inactivation of phosphorylase in hepatocytes by serotonergic mechanisms, and counter-regulation by atypical antipsychotic drugs. Diabetologia 50: 1743-1751
- Moore MC, Geho WB, Lautz M, Farmer B, Neal DW, et al. (2004) Portal infusion and glucose disposal in conscious dogs. Diabetes 53: 14–20.
- 16. Watanabe H, Akasaka D, Ogasawara H, Sato K, Miyake M, et al. (2010) Peripheral serotonin enhances lipid metabolism by accelerating bile acid turnover. Endocrinology 151: 4776–4786.
- 17. Bergman EN, Katz ML, Kaufman CF (1970) Quantitative aspects of hepatic and portal glucose metabolism and turnover in sheep. Am J Physiol 219: 785-
- 18. Bergman EN, Reulein SS, Corlett RE (1989) Effects of obesity on insulin ensitivity and responsiveness in sheep. Am J Physiol 257: 772-781
- 19. Brockman RP (1983) Effects of insulin and glucose on the production and utilization of glucose in sheep (Ovis aries). Comp Biochem Physiol A Mol Integr Physiol 74: 681-685.
- Seal CJ, Reynolds CK (1993) Nutritional implications of gastrointestinal and liver metabolism in ruminants. Nutr Res Rev 6: 185-208.
- 21. Jarrett IG, Potter BJ (1953) Insulin tolerance and hypoglycaemic convulsions in sheep. Aust J Exp Biol Med Sci 31: 311-318.
- 22. Robertson JP, Faulkner A, Vernon RG (1982) Regulation of glycolysis and fatty acid synthesis from glucose in sheep adipose tissue. Biochem J 206: 577-586.
- Smith SB, Crouse JD (1984) Relative contributions of acetate, lactate and glucose to lipogenesis in bovine intramuscular and subcutaneous adipose tissue. J Nutr 114: 792-800.
- Smith SB, Prior RL (1986) Comparisons of lipogenesis and glucose metabolism between ovine and bovine adipose tissues. J Nutr 116: 1279-1286.
- Vernon RG (1980) Lipid metabolism in the adipose tissue of ruminant animals Prog Lipid Res 19: 23-106.
- 26. Fukumori R, Sugino T, Hasegawa Y, Kojima M, Kangawa K, et al. (2011) Plasma ghrelin concentration is decreased by short chain fatty acids in wethers. Domest Anim Endocrinol 41: 50-55.
- 27. McCowen KC, Maykel JA, Bistrian BR, Ling PR (2002) Circulating ghrelin concentrations are lowered by intravenous glucose or hyperinsulinemic euglycemic conditions in rodents. J Endocrinol 175: R7-11.

Author Contributions

Conceived and designed the experiments: HW HS HK HA. Performed the experiments: HW RS TN HT YT K.Sumiyoshi K.Sato XC NO SI DH NS. Analyzed the data: HW HS SO KW HA. Contributed reagents/ materials/analysis tools: HW HA. Wrote the paper: HW MR HA.

- 28. Shiiya T, Nakazato M, Mizuta M, Date Y, Mondal MS, et al. (2002) Plasma ghrelin levels in lean and obese humans and the effect of glucose on ghrelin secretion. J Clin Endocrinol Metab 87: 240–244.
- 29. Itoh F, Komatsu T, Kushibiki S, Hodate K (2006) Effects of ghrelin injection on plasma concentrations of glucose, pancreatic hormones and cortisol in Holstein dairy cattle. Comp Biochem Physiol A Mol Integr Physiol 143: 97-102.
- Gershon MD (1999) Review article: serotonin roles played by 5-hydroxytryp-tamine in the physiology of the bowel. Aliment Pharmacol Ther 13: 15–30.
- 31. Kidd M, Modlin IM, Gustafsson BI, Drozdov I, Hauso O, et al. (2008) Luminal regulation of normal and neoplastic human EC cell serotonin release is mediated by bile salts, amines tastants, and olfactants. Am J Physiol Gastrointest Liver Physiol 295: G260-272
- 32. Peregrin A, Ahlman H, Jodal M, Lundgren O (1999) Involvement of serotonin and calcium channels in the intestinal fluid secretion evoked by bile salt and cholera toxin. Br J Pharmacol 127: 887-894.
- 33. Racke K, Reimann A, Schworer H, Kilbinger H (1996) Regulation of 5-HT release from enterochromaffin cells, Behav Brain Res 73: 83-87.
- 34. Zhu J, Wu X, Owvang C, Li Y (2001) Intestinal serotonin acts as a paracrine substance to mediate vagal signal transmission evoked by luminal factors in the at. J Physiol 530: 431-442.
- Schneider TJ, Struijk PC, Lotgering FK, Wallenburg HC (1996) Placental transfer and maternal and fetal hemodynamic effects of ketanserin in the pregnant ewe. Eur J Obstet Gynecol Reprod Biol 68: 179-184.
- 36. Thomas GB, Cummins JT, Clarke IJ (1987) Secretion of prolactin in response to serotonin requires an intact hypothalamo-pituitary axis in the ewe. Neurosci Lett
- 37. Zinner MJ, Kasher F, Jaffe BM (1983) The hemodynamic effects of intravenous
- infusions of serotonin in conscious dogs. J Surg Res 34: 171–178.

 38. Centurión D, Ortiz MI, Saxena PR, Villalón CM (2002) The atypical 5-HT2 receptor mediating tachycardia in pithed rats: pharmacological correlation with the 5-HT2A receptor subtype. Br J Pharmacol 135: 1531-1539.
- 39. Säfsten B, Sjöblom M, Flemström G (2006) Serotonin increases protective duodenal bicarbonate secretion via enteric ganglia and a 5-HT4-dependent pathway. Scand J Gastroenterol 41: 1279-1289.
- Paulmann N, Grohmann M, Voigt JP, Bert B, Vowinckel J, et al. (2009) Intracellular serotonin modulates insulin secretion from pancreatic beta-cells by protein serotonylation. PLoS Biol 7: e1000229.
- 41. Zawalich WS, Tesz GJ, Zawalich KC (2004) Effects of prior 5-hydroxytryptamine exposure on rat islet insulin secretory and phospholipase C responses. Endocrine 23: 11-16.
- 42. Hajduch E, Rencurel F, Balendran A, Batty IH, Downes CP, et al. (1999) Serotonin (5-Hydroxytryptamine), a novel regulator of glucose transport in rat skeletal muscle. J Biol Chem 274: 13563–13568.
- 43. Moore MC, Kimura K, Shibata H, Honjoh T, Saito M, et al. (2005) Portal 5hydroxytryptophan infusion enhances glucose disposal in conscious dogs. Am J Physiol Endocrinol Metab 289: E225-31.
- 44. Forte TM, Cross CE, Gunther RA, Kramer GC (1983) Characterization of sheep lung lymph lipoproteins: chemical and physical properties. J Lipid Res 24: 1358-1367
- Kozat S, Denizhan V (2010) Glucose, lipid, and lipoprotein levels in sheep naturally infected with Fasciola hepatica. J Parasitol 96: 657–659.
- Nakata K, Taniguchi Y, Yoshioka N, Yoshida A, Inagawa H, et al. (2011) A mixture of Salacia oblonga extract and IP-PA1 reduces fasting plasma glucos (FPG) and low-density lipoprotein (LDL) cholesterol levels. Nutr Res Pract 5:
- 47. Watanabe H, Rose MT, Aso H (2011) Role of peripheral serotonin in glucose and lipid metabolism. Curr Opin Lipidol 22: 186–191.
 Bogach PG, Liashchenko PS (1976) The effect of serotonin on bile secretion in
- dugs. Fiziologicheskii zhurnal SSSR imeni I.M. Sechenova 62: 283-288.
- 49. Jang JH, Rickenbacher A, Humar B, Weber A, Raptis DA, et al. (2012) Serotonin protects mouse liver from cholestatic injury by decreasing bile salt pool after bile duct ligation. Hepatology 56: 209-218.
- 50. Fernández JI, Naranjo JA, Valverde A, Rueda A, Martínez-Victoria E, et al. (1996) Post-natal changes in biliary lipids in suckling goat kids. Br Vet J 152: 673-682
- 51. Hoyer D, Hannon JP, Martin GR (2002) Molecular, pharmacological and functional diversity of 5-HT receptors. Pharmacol Biochem Behav 71: 533-554.

Effect of Peripheral 5-HT on Glucose and Lipid Metabolism in Wether Sheep

vveti	ner Sheep				
ORIGINA	ALITY REPORT				
SIMILA	8% ARITY INDEX	8% INTERNET SOURCES	16% PUBLICATIONS	% STUDENT F	PAPERS
PRIMAR	RY SOURCES				
1	Advance Biology,	es in Experimenta 1996.	Il Medicine and	d	1%
2	docplaye				1%
3	www.plo Internet Sourc	sone.org _e			1%
4	hydroxyt feeding a	C. Neill, Steven J ryptamine and d- and sham drinkin d rat", Physiology	fenfluramine og in the gastric	n sham :-	1%
5	research Internet Sourc	repository.murdo	och.edu.au		1%
6	link.sprin				1%
7		hi, T "Different ı dial plasma ghre	•	els	<1%

induced by concentrate or timothy hay feeding in wethers", Domestic Animal Endocrinology, 200805

Publication

Jun Yamada, Yumi Sugimoto, Tomoko Yoshikawa, Ikuko Kimura, Kazuyoshi Horisaka. "The involvement of the peripheral 5-HT2A receptor in peripherally administered serotonininduced hyperglycemia in rats", Life Sciences, 1995

<1%

Publication

9 stoa.usp.br

<1%

Md. Sharif Shajib, Adriana Baranov, Waliul I. Khan. "Diverse Effects of Gut-Derived Serotonin in Intestinal Inflammation", ACS Chemical Neuroscience, 2017

<1%

Publication

C. J. Seal, C. K. Reynolds. "Nutritional Implications of Gastrointestinal and Liver Metabolism in Ruminants", Nutrition Research Reviews, 2007

<1%

Publication

Meyer Friedman, Sanford O. Byers, Fred Michaelis. "Bile Acid Content of Rat Bile and of Normal and Icteric Rat Plasma", American Journal of Physiology-Legacy Content, 1951

<1%

Philipp Stahl, Volker Ruppert, Ralph T. <1% 13 Schwarz, Thomas Meyer. "Trypanosoma cruzi Evades the Protective Role of Interferon-Gamma-Signaling in Parasite-Infected Cells", PLoS ONE, 2014 Publication Peter Back. "Phenobarbital-induced alterations <1% of bile acid metabolism in cases of intrahepatic cholestasis", Klinische Wochenschrift, 06/1982 Publication hdl.handle.net <1% Internet Source Alsip, N.L.. "Serotonin-induced dilation of small 16 arterioles is not mediated via endotheliumderived relaxing factor in skeletal muscle", European Journal of Pharmacology, 19921215 Publication P. Pazzi. "Serum bile acids in patients with liver <1% 17 failure supported with a bioartificial liver", Alimentary Pharmacology and Therapeutics, 8/2002 Publication Medical Science Symposia Series, 1993. 18 Publication

- 20
- U. Haus. "Spectrum of use and tolerability of 5-HT3 receptor antagonists", Scandinavian Journal of Rheumatology, 6/2004

<1%

Publication

- 21
- S. Oda, M. Ikuta, T. Kuhara, A. Ohneda, Y. Sasaki. "Insulin and glucagon secretion in goats (Capra hircus Linnæus) exposed to cold", Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology, 1995

<1%

Publication

22

Hiroki Ikeda, Tomoki Shibuya, Shunsuke Imanishi, Hisashi Aso, Manabu Nishiyama, Yoshinori Kanayama. "Dynamic Metabolic Regulation by a Chromosome Segment from a Wild Relative During Fruit Development in a Tomato Introgression Line, IL8-3", Plant and Cell Physiology, 2016 <1%

Publication

23

Kubátová, Anna, and Tamara Fedorova. "Saliva Crystallization Occurs in Female Bornean Orangutans (Pongo pygmaeus): Could It Be a New Option for Monitoring of Menstrual Cycle in Captive Great Apes?", PLoS ONE, 2016.

<1%

Publication

24	Anna Kubátová, Tamara Fedorova. "Saliva Crystallization Occurs in Female Bornean Orangutans (Pongo pygmaeus): Could It Be a New Option for Monitoring of Menstrual Cycle in Captive Great Apes?", PLOS ONE, 2016 Publication	<1%
25	www.biochemj.org Internet Source	<1%
26	www.ncbi.nlm.nih.gov Internet Source	<1%
27	Masae Shiyomi, Shigeo Takahashi, Jin Yoshimura, Taisuke Yasuda, Michio Tsutsumi, Mikinori Tsuiki, Yoshimichi Hori. "Spatial heterogeneity in a grassland community: Use of power law", Ecological Research, 2001 Publication	<1%
28	"Pharmacology of Smooth Muscle", Springer Science and Business Media LLC, 1994 Publication	<1%
29	edoc.mdc-berlin.de Internet Source	<1%
30	Chun-yu Huang, Jing-jing Zhao, Lin Lv, Yi-bing Chen et al. "Decreased Expression of AZGP1 Is Associated with Poor Prognosis in Primary Gastric Cancer", PLoS ONE, 2013 Publication	<1%

31	preview- nutritionandmetabolism.biomedcentral.com Internet Source	<1%
32	www.frontiersin.org Internet Source	<1%
33	Namba, H "Transforming growth factor-@a changes firing properties of developing neocortical GABAergic neurons by down-regulation of voltage-gated potassium currents", Neuroscience, 2003 Publication	<1%
34	jcancer.org Internet Source	<1%
35	shareok.org Internet Source	<1%
36	Jimena Laporta, Tonia L. Peters, Kathryn E. Merriman, Chad M. Vezina, Laura L. Hernandez. "Serotonin (5-HT) Affects Expression of Liver Metabolic Enzymes and Mammary Gland Glucose Transporters during the Transition from Pregnancy to Lactation", PLoS ONE, 2013 Publication	<1%
37	E. N. Bergman, S. S. Reulein, R. E. Corlett.	<1%

"Effects of obesity on insulin sensitivity and responsiveness in sheep", American Journal of

Physiology-Endocrinology and Metabolism, 1989

Publication

Ram Jagannathan, Azizi Seixas, David St-Jules, Lakshmanan Jagannathan et al. "Systems Biology Genetic Approach Identifies Serotonin Pathway as a Possible Target for Obstructive Sleep Apnea: Results from a Literature Search Review", Sleep Disorders, 2017

Publication

programinplacebostudies.org

<1%

<1%

A G Dayer. "Excess of serotonin affects embryonic interneuron migration through activation of the serotonin receptor 6", Molecular Psychiatry, 07/29/2008

<1%

Publication

r. mitsui. "Neural and non-neural mediation of propionate-induced contractile responses in the rat distal colon", Neurogastroenterology and Motility, 8/2005

<1%

Publication

S Rozenblit-Susan, N Chapnik, O Froy.
"Serotonin prevents differentiation into brown adipocytes and induces transdifferentiation into white adipocytes", International Journal of Obesity, 2017

<1%

43

Yumi Sugimoto, Jun Yamada, Tomoko Yoshikawa, Toshiko Noma, Kazuyoshi Horisaka. "Effects of peripheral 5-HT2 and 5-HT3 receptor agonists on food intake in food-deprived and 2deoxy-d-glucose-treated rats", European Journal of Pharmacology, 1996

<1%

Publication

44

www.wfsbp.org

Internet Source

<1%

45

Pierre Blier. "Electrophysiological assessment of putative antagonists of 5-hydroxytryptamine receptors: a single-cell study in the rat dorsal raphe nucleus", Canadian Journal of Physiology and Pharmacology, 02/1989

<1%

Publication

46

repub.eur.nl

Internet Source

<1%

47

Susan M. Butler, Anne Faulkner, Victor A. Zammit, Richard G. Vernon. "Fatty acid metabolism of the perfused caudate lobe from livers of fed and fasted non-pregnant and fasted late pregnant ewes", Comparative Biochemistry and Physiology Part B: Comparative Biochemistry, 1988

Publication

48	www.ircbt.net Internet Source	<1%
49	jeb.biologists.org Internet Source	<1%
50	Daniel D. Lam, Lora K. Heisler. "Serotonin and energy balance: molecular mechanisms and implications for type 2 diabetes", Expert Reviews in Molecular Medicine, 2007 Publication	<1%
51	journalofanimalscience.org Internet Source	<1%
52	Itoh, F "Effects of ghrelin injection on plasma concentrations of glucose, pancreatic hormones and cortisol in Holstein dairy cattle", Comparative Biochemistry and Physiology, Part A, 200601 Publication	<1%
53	www.clinsci.org Internet Source	<1%
54	www.plosbiology.org Internet Source	<1%
55	Kazuhito Sakamoto, Tokushi Komatsu, Takuya Kobayashi, Michael T Rose, Hisashi Aso, Akihiko Hagino, Yoshiaki Obara. "Growth hormone acts on the synthesis and secretion of	<1%

α-casein in bovine mammary epithelial cells", Journal of Dairy Research, 2005

Publication

56	Antonio Pisani. "Endogenous Serotonin Excites Striatal Cholinergic Interneurons via the Activation of 5-HT 2C, 5-HT6, and 5-HT7 Serotonin Receptors: Implications for Extrapyramidal Side Effects of Serotonin Reuptake Inhibitors", Neuropsychopharmacology, 08/2007 Publication	<1%
57	bioone.org Internet Source	<1%
58	Christina S Chao. "Ghrelin, insulin, and the pancreatic islet", Current Opinion in Endocrinology & Diabetes, 04/2004 Publication	<1%
59	J. Balcells, C. J. Seal, D. S. Parker. "Effect of intravenous glucose infusion on metabolism of portal-drained viscera in sheep fed a cereal/straw-based diet2", Journal of Animal Science, 1995 Publication	<1%
60	Progress in Experimental Cardiology, 2003. Publication	<1%

ajpendo.physiology.org

<1%

62	Hsieh, J.T "The Activation of Peripheral 5- HT1A Receptors Can Inhibit Seminal Vesicle Contraction: An In Vivo Animal Study", Urology, 201108 Publication	<1%
63	Alan F. Hofmann. "Bile Acids and the Enterohepatic Circulation", The Liver, 11/13/2009 Publication	<1%
64	www.physiology.org Internet Source	<1%
65	Kanarek, R.B "Peripheral serotonin administration selectively reduces fat intake in rats", Pharmacology, Biochemistry and Behavior, 198809 Publication	<1%
66	Wagner, C "Intraduodenal serotonin elicits non-propagating spike potentials in the small intestine of the rat", Comparative Biochemistry and Physiology, Part A, 200311 Publication	<1%
67	I. Kaji. "Effects of luminal thymol on epithelial transport in human and rat colon", AJP Gastrointestinal and Liver Physiology, 03/03/2011 Publication	<1%

- 68
- M.G. Marrero, B. Dado-Senn, S.L. Field, D.R. da 69 Silva, A.L. Skibiel, J. Laporta. "Increasing serotonin bioavailability in preweaned dairy calves impacts hematology, growth, and behavior", Domestic Animal Endocrinology, 2019

Publication

www.nature.com 70 Internet Source

<1_%

Bin Li, Yun-Wen Zheng, Yuuki Sano, Hideki 71 Taniguchi. "Evidence for Mesenchymal-Epithelial Transition Associated with Mouse Hepatic Stem Cell Differentiation", PLoS ONE, 2011

Publication

Goyal, R. K., V. Elimban, Y.-J. Xu, H. 72 Kumamoto, N. Takeda, and N. S. Dhalla. "Mechanism of Sarpogrelate Action in Improving Cardiac Function in Diabetes", Journal of Cardiovascular Pharmacology and Therapeutics, 2011.

<1%

Publication

pdfs.semanticscholar.org Internet Source

Hiroya KADOKAWA, Katsuhiro AlKAWA, Koji KIMURA, Domique BLACHE, Ian H. WILLIAMS, Graeme B. MARTIN. "Links between De Novo Fatty Acid Synthesis and Leptin Secretion in Bovine Adipocytes", Journal of Veterinary Medical Science, 2007

<1%

Publication

Eiko Takishita. "Effect of Sarpogrelate
Hydrochloride, A 5-HT2 Blocker, on Insulin
Resistance in Otsuka Long-Evans Tokushima
Fatty Rats (OLETF rats), A Type 2 Diabetic Rat
Model", Journal of Cardiovascular

<1%

Publication

Pharmacology, 02/2004

H. Kaess, W. Ehlers, A. Burkhardt. "Distribution of Electrolytes and Insulin Content in the Pancreas of Potassium Deficient Rats", Hormone and Metabolic Research, 2009

<1%

Publication

F. Carvalho, D. Macêdo, I. Bandeira, I.
Maldonado, L. Salles, M. Azevedo, M. Rocha
Jr., J. Fregoneze, E. De Castro-e-Silva. "
Central 5-HT Receptor Stimulation by m-CPBG
Increases Blood Glucose in Rats ", Hormone

and Metabolic Research, 2002

<1%

Publication

Johnson, "Anteroventral third ventricle lesions <1% attenuate pressor responses to serotonin in in anesthetized rats", Brain Research, 1996 Publication Fukumori, R.. "Plasma ghrelin concentration is <1% 79 decreased by short chain fatty acids in wethers", Domestic Animal Endocrinology, 201107 Publication Nils Paulmann. "Intracellular Serotonin <1% 80 Modulates Insulin Secretion from Pancreatic β-Cells by Protein Serotonylation", PLoS Biology, 10/27/2009 Publication <1% Tubio, R. I. C., J. Perez-Maceira, and M. 81 Aldegunde. "Homeostasis of glucose in the rainbow trout (Oncorhynchus mykiss Walbaum): the role of serotonin", Journal of Experimental Biology, 2010. Publication Marie Hagbom. "Rotavirus Stimulates Release <1% 82 of Serotonin (5-HT) from Human Enterochromaffin Cells and Activates Brain Structures Involved in Nausea and Vomiting", PLoS Pathogens, 07/14/2011 Publication

Exclude quotes On Exclude matches Off

Exclude bibliography On