

direct extraction of cells, or callus culture in plants. To be able to perform the extraction of necessary first cell suspension cultures. Cell suspension cultures, can be obtained by growing callus or crumbs from leaf mesophyll cells. It is possible to obtain cell suspension cultures from leaf mesophyll of which required several stages of cell isolation from leaf mesophyll tissue, purification cells and cell culture. At the time of cell isolation, there are several factors that affect success of cell isolation. Factors, among others, the source of explant, type of explant, concentration of enzyme pectinase, osmoticum, long incubation time and method of incubation and pH of the medium.

At this stage of cell suspension cultures, often a variety of media both on the composition of the media, culture and explant own environment to secondary metabolite production [4]. The concentration of sugar in the medium range growth of 1-10% for secondary metabolite production. The goal of this research was to determine the effect of sucrose concentration and the concentration of leaf *C. asiatica* on the growth of cell suspension culture using R-10 which can generate a number of cells and cell viability for leaf mesophyll cell suspension culture of *C. asiatica*. The second aim is to investigate the effect of different concentrations of sucrose on the growth of cell suspension culture and the formation of compounds asiaticoside.

RESEARCH METHODS

Materials Research of explant

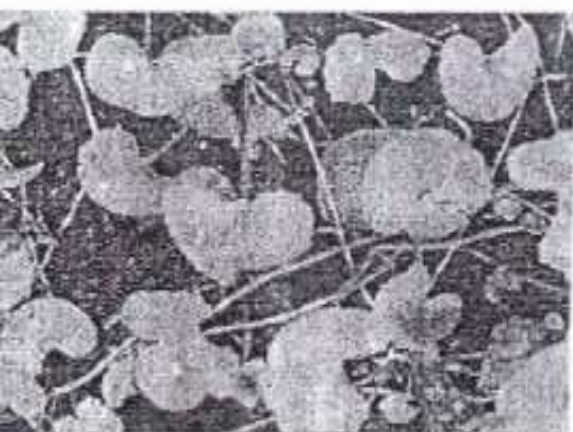


Fig.1. *Centella asiatica* L. Urban

Reagents and chemicals

Macerozyme R-10, bayclin, 96% alcohol, sterile distilled water

Media

MS medium, 2,2-dimethyl-1,3-bis(vinylpyrrolidone) from Sigma, sorbitol,

2-N-Morpholino ethane sulphonic acid (MES), macerozyme R-10 from Yakult Co. HONSA., Japan.

4. Purification medium

KNO₃, Ca (NO₃)₂, sorbitol and MES

5. Cells suspension culture medium

For cell suspension culture used Murashige and Skoog medium or MS medium, plant growth regulator (Indole 3-Acetic Acid) and BAP (Benzylaminopurine)

6. Fensosafranin 0.2% solution of the sigma brand for observation of cell viability

C. Research Tools

1. **sterilization** ; cell isolation and purification of cells and cell culture were as follows: laminar air flow cabinet, UV lamp equipped, autoclave, incubation space, light microscope, dissecting set, magnetic stirrer, analytical scales, laboratory equipment, centrifuges and conic tube, paper filter 0.22 μ milipore from Whatman, nylon mesh size 100, the light microscope, photomicroskop, pH paper, glass slides, umbrellas, micrometers, hemocytometer, counting chamber, aluminum foil, shakers, oven, pipette, holder, and incubators.

2. chemical analysis

Glass tools for the extraction of cells, tools for thin layer chromatography, UV lamp 254 nm and 265 nm, Shimadzu brand, bath water, and eksikator.

Procedure

At this stage of isolation and purification of mesophyll cells, the experimental design used was completely randomized design factorial. The first factor is the sequence of leaf: leaf sequence-1 (D1), 2nd (D2) and 3rd (D3). Macerozyme second factor is the concentration of R-10 is E0 = 0% (0g), E1 = 0, 1% (1g / l), E2 = 0.3% (2g / l) and E3 = 0.3% (3g / l). Each combination of treatments with three replicates. The parameters measured were the number of cells and cell viability. The data obtained were analyzed by ANOVA Duncan Test's Multiple Range Test (DMRT) at 5% confidence level. While at this stage of cell suspension cultures tested the addition of sucrose treatment on production medium with a concentration of 0%, 2.5% and 7.5%. To measure the growth of both cell suspension cultures in growth medium and the medium of production is done by measuring packed cell volume (PCV) in a certain time interval.

RESULT AND DISCUSSION

1. Obtaining the number and viability of cells