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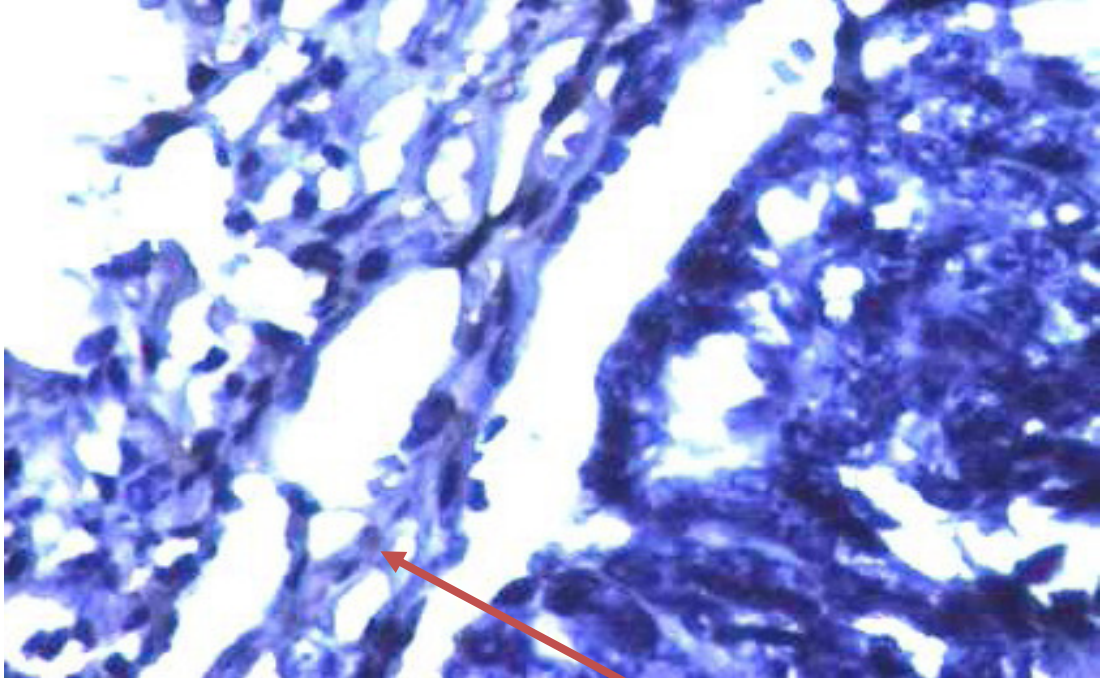
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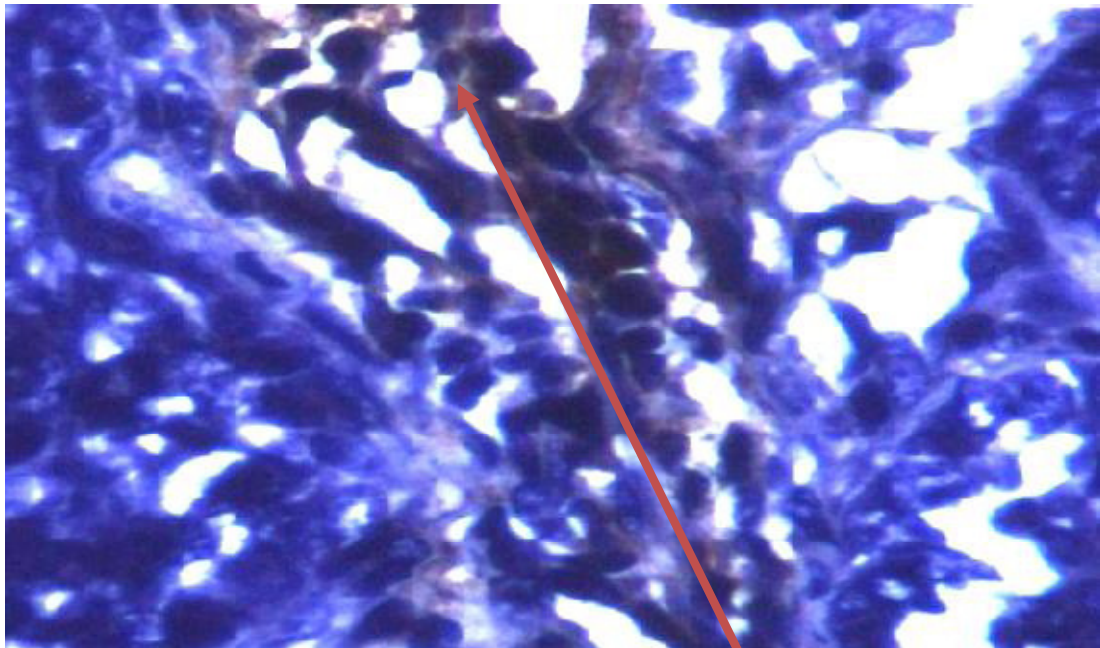
LAMPIRAN 1

HASIL PEMERIKSAAN Sel T CD4⁺



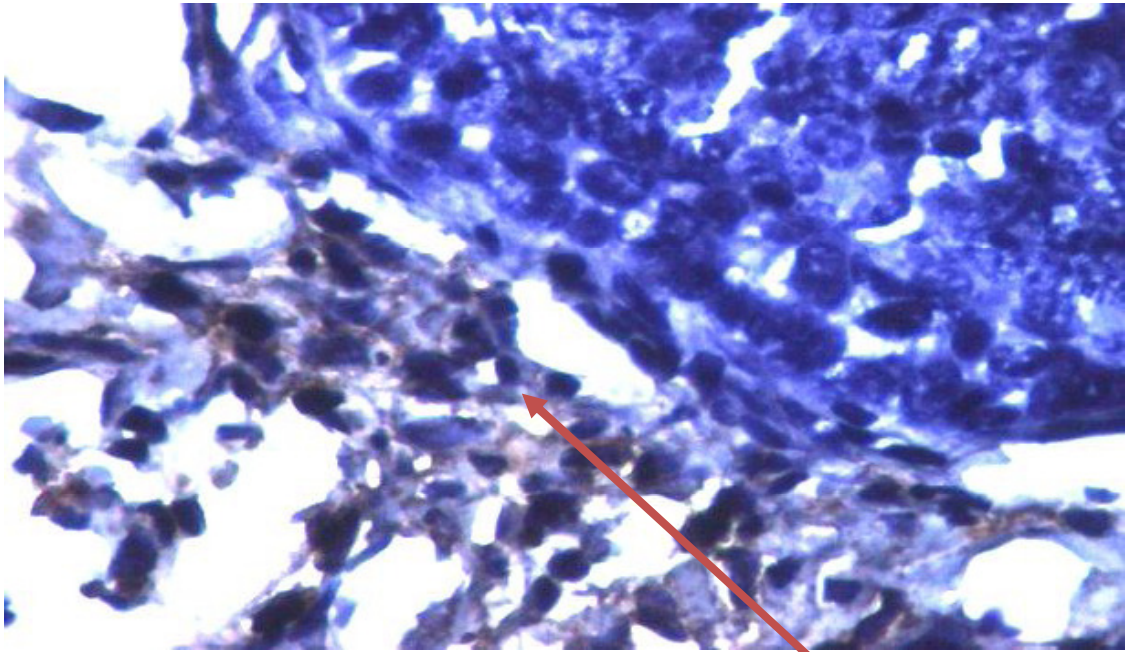
KONTROL

Gb 1. Ada 1 sel T CD4⁺
pada 1 lap pandang



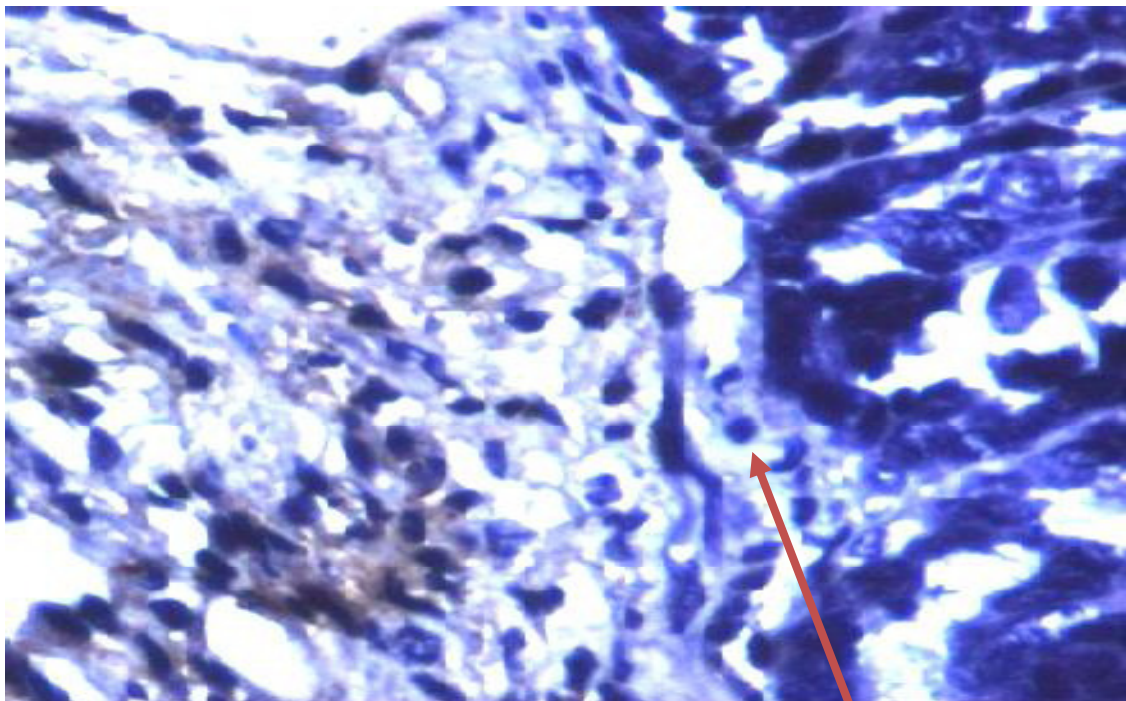
Cyclophosphamide (P1)

Gb 2. Ada 2 sel T CD4⁺
pada 1 lap pandang



Transfer Factor (P2)

GB 3. Ada 2 sel T CD4⁺
pada 1 lap pandang



Kombinasi (P3)

Gb 4. Ada 5 sel T CD4⁺
pada 1 lap pandang

LAMPIRAN 2

Data

Kelompok	Lap 1	Lap 2	Lap 3	Lap 4	Lap 5	Jumlah
Kontrol	1	2	2	1	2	8
	0	1	2	1	2	6
	1	1	1	2	2	7
	2	1	1	1	1	6
	2	2	1	2	1	8
P1 (Cyclo)	4	2	2	2	3	13
	3	2	3	2	4	14
	4	4	3	3	4	18
	3	3	3	3	4	13
	2	4	2	4	3	15
P2 (TF)	3	3	2	3	3	14
	3	4	2	2	4	15
	3	3	2	4	2	14
	3	4	3	2	4	16
	4	2	4	3	3	16
P3 (Komb)	4	3	4	4	2	17
	4	4	5	2	5	20
	5	4	5	4	2	20
	5	5	3	5	4	22
	3	6	6	4	2	21

LAMPIRAN 3

ANALISIS STATISTIK

Descriptives

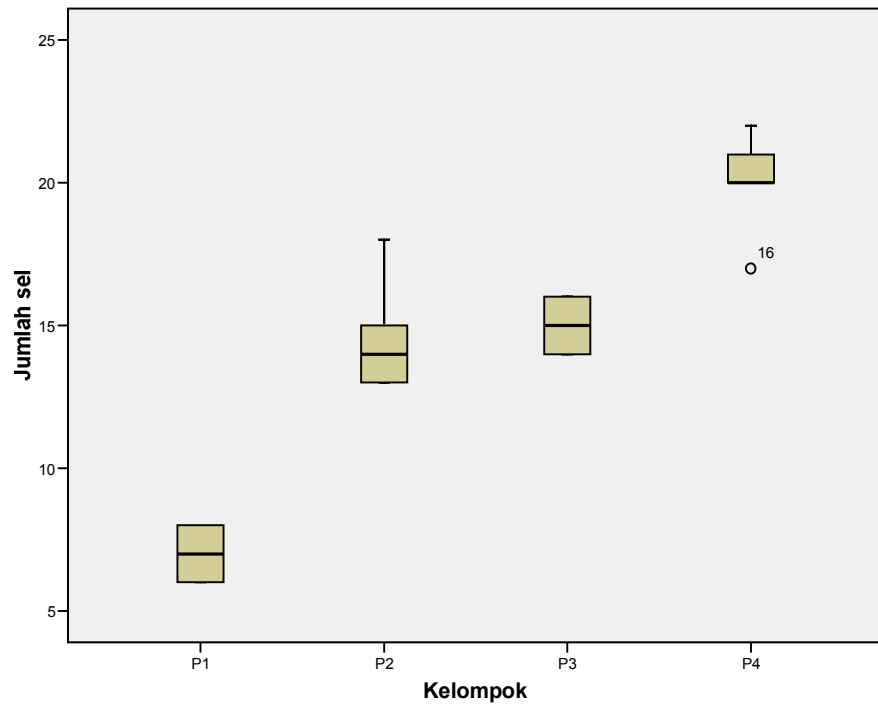
Kelompok		Statistic	Std. Error			
Jumlah sel	P1	Mean	7.00	.447		
		95% Confidence Interval for Mean	Lower Bound	5.76		
			Upper Bound	8.24		
		5% Trimmed Mean	7.00			
		Median	7.00			
		Variance	1.000			
		Std. Deviation	1.000			
		Minimum	6			
		Maximum	8			
		Range	2			
		Interquartile Range	2			
		Skewness	.000	.913		
		Kurtosis	-3.000	2.000		
		P2	P2	Mean	14.60	.927
				95% Confidence Interval for Mean	Lower Bound	12.03
Upper Bound	17.17					
5% Trimmed Mean	14.50					
Median	14.00					
Variance	4.300					
Std. Deviation	2.074					
Minimum	13					
Maximum	18					
Range	5					
Interquartile Range	4					
Skewness	1.447			.913		
Kurtosis	1.931			2.000		
P3	P3			Mean	15.00	.447
				95% Confidence Interval for Mean	Lower Bound	13.76
		Upper Bound	16.24			
		5% Trimmed Mean	15.00			
		Median	15.00			
		Variance	1.000			
		Std. Deviation	1.000			
		Minimum	14			
		Maximum	16			
		Range	2			
		Interquartile Range	2			
		Skewness	.000	.913		

P4	Kurtosis		-3.000	2.000
	Mean		20.00	.837
	95% Confidence Interval for Mean	Lower Bound	17.68	
		Upper Bound	22.32	
	5% Trimmed Mean		20.06	
	Median		20.00	
	Variance		3.500	
	Std. Deviation		1.871	
	Minimum		17	
	Maximum		22	
	Range		5	
	Interquartile Range		3	
	Skewness		-1.145	.913
	Kurtosis		2.000	2.000

Tests of Normality

kelompok		Kolmogorov-Smirnov ^a			Shapiro-Wilk			
		Statistic	Df	Sig.	Statistic	df	Sig.	
data	kontrol	.241	5	.200	kontrol	.231	5	.119
	cyclo	.224	5	.200	cyclo	.221	5	.171
	TF	.241	5	.200	TF	.273	5	.119
	Komb	.300	5	.161	Komb	.908	5	.453

a. Lilliefors Significance Correction



Oneway

Test of Homogeneity of Variances

Jumlah sel

Levene Statistic	df1	df2	Sig.
.692	3	16	.570

ANOVA

Jumlah sel

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	431.350	3	143.783	58.687	.000
Within Groups	39.200	16	2.450		
Total	470.550	19			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Jumlah sel

Tukey HSD

(I) Kelompok	(J) Kelompok	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
P1	P2	-7.600*	.990	.000	-10.43	-4.77
	P3	-8.000*	.990	.000	-10.83	-5.17
	P4	-13.000*	.990	.000	-15.83	-10.17
P2	P1	7.600*	.990	.000	4.77	10.43
	P3	-.400	.990	.977	-3.23	2.43
	P4	-5.400*	.990	.000	-8.23	-2.57
P3	P1	8.000*	.990	.000	5.17	10.83
	P2	.400	.990	.977	-2.43	3.23
	P4	-5.000*	.990	.001	-7.83	-2.17
P4	P1	13.000*	.990	.000	10.17	15.83
	P2	5.400*	.990	.000	2.57	8.23
	P3	5.000*	.990	.001	2.17	7.83

*. The mean difference is significant at the .05 level.

LAMPIRAN 4

1. jaringan tumor pada mencit.
 1. Alkohol Bahan transplantasi 70 %
 2. Larutan Garam fisiologik
 3. Es batu
 4. Mencit donor bertumor
 5. Mencit resipien

2. Bahan untuk pemeriksaan pengecatan Sel T CD4⁺.
 1. Polinized slide
 2. Xylol
 3. Alkohol absolut
 4. Tris Hcl (pH 7,6)
 5. Periodic acid 0,5 %
 6. 0,3 % H₂O₂ dalam metanol
 7. Blok 1 % BSA – PBS
 8. Biotinylated Rabbit anti Mouse
 9. Ab Sekunder
 10. DAB
 11. Sel T CD4⁺ mouse Ab
 12. Hematoksilin
 13. Entelan

LAMPIRAN 5

Prosedur transplantasi tumor

1. Mencit donor dimatikan dengan eter, kemudian diletakkan terlentang pada tatakan / alas fiksasi dan keempat kakinya difiksasi dengan jarum.
2. Kulit dibagian yang bertumor diusap dengan alkohol 70 %, kemudian dibuat sayatan dengan gunting lurus, untuk mengeluarkan tumor.
3. Tumor diletakkan di cawan petri kecil yang telah terlebih dahulu dicuci dengan garam fisiologis dan diletakkan diatas es.
4. Amati bentuk dan keadaan tumor, kemudian ambil/potong jaringan tumor yang masih baik yaitu bagian yang tanpa nekrosis (biasanya di daerah tepi jika tumor besar) sebanyak kira-kira yang dapat menghasilkan bubur tumor paling sedikit 1 ml dan taruh dicawan petri kecil lainnya. Bersihkan dari jaringan ikat (simpai), jaringan nekrotik dan darah, kemudian cacah/potong-potong sampai halus dengan gunting hingga akhirnya terbentuk “bubur tumor” yang partikelnya dapat melewati jarum trokar. Tambahkan garam fisiologis lebih kurang sama banyak dengan volume tumor.
5. Bubur tumor disuntikkan subkutan di aksila kanan mencit dengan dosis 0,2 ml.
6. Sisa tumor yang padat dimasukkan ke dalam botol formalin untuk dibuat sediaan mikroskopik histopatologi.
7. Masing-masing mencit diberi nomor ditelinganya (lihat bagan) dan dimasukkan ke dalam kandang berbeda yang diberi label berisi : jenis kelompok perlakuan, tanggal transplantasi .

LAMPIRAN 6

Prosedur pembuatan preparat blok paraffin

a. Fiksasi

Potongan adenokarsinoma dimasukkan dalam larutan formalin buffer (larutan formalin 10% dalam buffer Natrium asetat sampai mencapai pH 7,0). Waktu fiksasi jaringan 18-24 jam. Setelah fiksasi selesai, jaringan dimasukkan dalam larutan aquades selama 1 jam untuk proses penghilangan larutan fiksasi.

b. Dehidrasi

Potongan adenokarsinoma dimasukkan dalam alkohol konsentrasi bertingkat. Jaringan menjadi lebih jernih dan transparan. Jaringan kemudian dimasukkan dalam larutan alkohol-xylool selama 1 jam dan kemudian larutan xylool murni selama 2x2 jam.

c. Impregnasi

Jaringan dimasukkan dalam paraffin cair selama 2x2 jam.

d. Embedding

Jaringan ditanam dalam paraffin padat yang mempunyai titik lebur 56-58°C, ditunggu sampai paraffin padat. Jaringan dalam paraffin dipotong setebal 4 mikron dengan mikrotom. Potongan jaringan ditempelkan pada kaca obyek yang sebelumnya telah diolesi polilisin sebagai perekat. Jaringan pada kaca obyek dipanaskan dalam inkubator suhu 56-58°C sampai paraffin mencair.

LAMPIRAN 7

Prosedur Pengecatan Sel T CD4⁺

1. Fiksasi dengan Polinized slide 3 menit
2. Deparafinisasi dengan Xylol dalam inkubator semalam
3. Rehidrasi dengan Alkohol absolut, kemudian dengan air murni
4. Cuci dengan 50 M Tris – HCl; selama 3 menit
5. Periodic Acid 0,5 % selama 10 menit
6. 0,3 % H₂O₂ dalam methanol selama 20 menit
7. Cuci dengan 50 M Tris – HCl selama 5 menit
8. Blok dengan 1 % BSA – PBS selama 30 menit
9. Inkubasi primer Ab CD4⁺ selama 1 jam
10. Cuci dengan 50 M Tris HCl selama 5 menit, 2 X
11. Biotynilated Rabbit anti Mouse selama 1 jam
12. Cuci dengan 50 M Tris HCl selama 5 menit, 2 X
13. Inkubasi Ab Sekunder selama 30 menit
14. Cuci dengan 50 M Tris HCl selama 5 menit, 2 X
15. DAB dalam Tris _ HCl kurang dari 10 menit
16. Couterstain Hematoksilin
17. Dehidrasi
18. Cleaning
19. Mountain dengan Entelan